

# Immunofluorescence Staining

Protocol for preparation cell suspension for cell sorting

Flow Cytometry Core Facility

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## **Procedure**

1. Add  $0.5 \times 10^6$  -  $1 \times 10^6$  cells to polystyrene tubes.
2. Spin down cells at  $300 \times g$  for 10 minutes at  $4^{\circ}\text{C}$ .
3. Remove supernatant.
4. Resuspend cell pellet in 50-100  $\mu\text{l}$  of staining buffer containing conjugated monoclonal antibodies each at  $1\mu\text{g} \times 10^6$  cells (or use dilution determined by titration for optimal staining). If using biotin conjugate, always stain with this antibody first and by itself.
5. Incubate on ice 20-30 minutes (Keep away from the light).
6. Wash once by adding 1ml 1X PBS / 1% FCS
7. Spin down cells at  $300 \times g$  for 10 minutes at  $4^{\circ}\text{C}$ .
8. If using biotin conjugate, resuspend cell pellet in 50 $\mu\text{l}$  of staining buffer (1X PBS / 1% FCS) containing streptavidin conjugated dye plus other directly conjugated antibodies, repeat steps 6 through 8.
9. Resuspend cell pellet in Hank's Buffered saline, PBS or [fixative](#) (analyze unfixed cells on the same day). Adjust concentration of cell suspension to at least  $10^6$  cells/ml
10. add Propidium Iodide or other available viability dye to unfixed stained cells to discriminate live from dead cells
11. An unstained sample or negative Ig control and single stained samples for each conjugated dye are also necessary for setting up the flow cytometer.

## **Reagents**

### ***Staining buffer***

1X PBS / 1% FCS or 1%BSA

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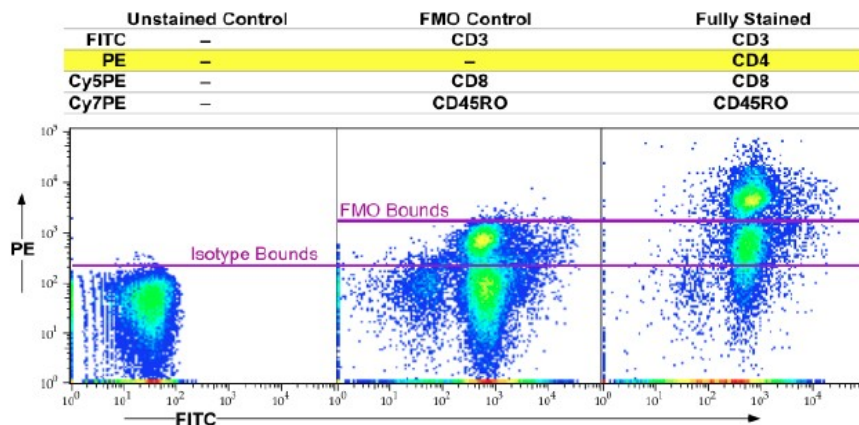
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## Controls that are necessary:

- Unstained cells: show the background or autofluorescence of the system.
- Isotype control: to identify non-specific binding. Isotype controls are good staining controls to identify potential problems in staining, particularly if a primary and secondary antibody are used.
- Sample with secondary reagent added only or similarly labeled non-specific primary antibody, ideally of the same isotype as the specific antibody you will be using
- Negatives and Positives control for each different primary antibody and each different cell population used in your experiment, when possible.
- FMO (Fluorescence Minus One) control: the cells were stained with all reagents except one. FMO control is essential when antigen expression is low or variable.

The following figure shows how a gate on an unstained or isotype would be set incorrectly as compared to an FMO control. Furthermore, the second panel demonstrates how FMO controls can resolve gating boundaries even with improper compensation:



## References

<https://www.amrepflow.org.au/public/documents/FCMcontrolsJTrotter2006.pdf>